3. Puromycin kill curve

Optimization of puromycin selection condition

To generate a fully transduced population of cells it is important to determine the minimum amount of puromycin required to eliminate non-transduced cells. This is accomplished by performing a kill curve to determine the optimal puromycin concentration needed to eliminate non-transduced cells. A kill curve is a dose-response experiment where the cells are subjected to increasing amounts of antibiotic to determine the minimum antibiotic concentration needed to kill all the cells over the course of 2 to 7 days. For puromycin, the optimal concentration is the lowest concentration that kills 100% of non-transduced cells and shows maximal survival of transduced cells in 48-72 hours. The optimal concentration is cell type dependent. Performing a kill curve is recommended with each new cell type or when a new selection antibiotic or different lot of selection antibiotic is used.

Puromycin Kill Curve

- 1. Plate cells at appropriate density in 100ul complete growth medium per well in a 96-well clear bottom tissue culture plate. The recommended cell density for most cell types is 30-50% confluence at the day of transduction. Incubate at 37°C overnight.
- 2. Replace media with 45ul media containing 4ug-8ug/ml polybrene(depend on cell type).
- 3. Infect the cells with shRNA control virus (C6-4) at MOI 2 and MOI 6:

| Virus MOI | | | | | | | | | | | | |
|-----------|--|---|---|---|---|---|---|---|---|---|---|--|
| A | | | | | | | | | | | | |
| В | | | | | | | | | | | | |
| C | | | | | | | | | | | | |
| D | | | | | | | | | | | | |
| E | | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | |
| F | | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | |
| G | | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | |
| Н | | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | 6 | |

- 4. Spin the plates at 1100g at 30°C for 15-30min. Incubate at 37°C overnight.
- 5. Replace the media from the infected cell plates and add 100ul media containing various puromycin concentrations as shown below. Incubate at 37°C for 48-72 hours.

| Puromycin concentration, ug/mL | | | | | | | | | | | | |
|--------------------------------|--|---|-----|---|---|---|---|---|---|---|---|--|
| A | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| В | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| C | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| D | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| E | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| F | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| G | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |
| Н | | 0 | 0.5 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | |

- 6. Remove the puromycin containing media from the cell plates.
- 7. Add 100ul of phenol red free DMEM medium containing 10% MTS. Incubate at 37°C for 0.5-3hrs.
- 8. Record the absorbance at 490 nm using a 96-well plate reader.
- 9. Determine the optimal puromycin concentration by choosing the minimum concentration of puromycin that causes complete cell death in non-transduced cells and maximal viability in transduced cells.